



# Variation of soil organic carbon, stable isotopes and soil quality indicators across an eroding-deposition catena in an historical Spanish olive orchard

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**Abstract.** This study compares the distribution of bulk soil organic carbon (SOC also reported as  $C_{org}$ ), its fractions  
15 (unprotected, physical, chemical and biochemically protected), available P ( $P_{avail}$ ), organic nitrogen ( $N_{org}$ ) and stable isotopes ( $\delta^{15}N$  and  $\delta^{13}C$ ) signatures at four soil depths (0–10, 10–20, 20–30, 30–40 cm) between a nearby forested reference area and an historical olive orchard (established in 1856) located in Southern Spain. In addition, these soil properties, as well as water stable aggregates ( $W_{sagg}$ ) were contrasted at eroding and deposition areas within the olive orchard, previously determined using  $^{137}Cs$ . Results highlight a significant depletion of SOC stock in the olive orchard as compared to the forested area,  
20 approximately 120 vs. 55 t C ha<sup>-1</sup> at the top 40 cm of soil respectively, being severe in the case of unprotected carbon fraction. Erosion and deposition within the old olive orchard created large differences in soil properties along a catena, resulting in higher  $C_{org}$ ,  $P_{avail}$  and  $N_{org}$  contents and  $\delta^{15}N$  at the deposition area and therefore defining two areas with a different soil quality status (degraded vs. non-degraded). Differences in  $\delta^{15}N$  at such different catena locations suggest that this isotopic signature has the potential for being used as an indicator of soil degradation magnitude, although additional  
25 studies would be required to confirm this finding. These overall results indicate that proper understanding of  $C_{org}$  content and soil quality in olive orchards require the consideration of the spatial variability induced by erosion/deposition processes for a convenient appraisal at farm scale.

## 1 Introduction

30 Research on soil organic carbon (SOC) and its dynamics has increased after the declaration of 4 per thousand program (Lal, 2015), which seeks to increase global soil organic matter stocks by 0.4 percent per year as a compensation for the global emissions of greenhouse gases by anthropogenic sources. Under this program special emphasis is given to combat soil



degradation, as it is a process which has a strong impact on the global carbon cycle because of the depletion of the SOC stock. For instance, in European agricultural soils, Lugato et al. (2016) reported that erosion-induced SOC fluxes were in the same order as the current gains from improved management and must be reduced to maintain soil health and productivity.

35 Lal (2003) estimated the global erosion-induced displacement of SOC at  $5.7 \text{ Pg C yr}^{-1}$ , and of that amount approximately 70 % is redistributed and redeposited over the landscape and the remaining 30% is transported by rivers into aquatic ecosystems. SOC is the most important indicator of soil quality (Rajan et al., 2010) and erosion-induced loss of SOC affects soil fertility on-site and the environment quality off-site (Lal, 2019). **However, the effects of soil erosion and the fate of the specific SOC fraction transported by erosion remains poorly understood.** Because of the agro-environmental impact of SOC

40 dynamics and SOC variability, more site and crop specific investigations are needed.

Olives are one of the most important crops in the Mediterranean region, where they cover approximately 9.7 Mha (FAOSTAT, 2019). Within the olive growing area there are regions where it becomes a dominant crop with a huge social and environmental significance, shaping the landscape of a great proportion of the territory. One of these regions is Andalusia. Located in Southern part of Spain, this is a region where olives cover approximately 17 % (Gómez et al., 2014)

45 and about 36 % of the agrarian utile territory. Olive cultivation has been linked to severe environmental issues including the acceleration of erosion and soil degradation (e.g. Beaufoy, 2001, Scheidel and Krausmann, 2011). In fact, soil degradation is common in olive orchards as they have been traditionally cultivated under rainfed conditions on sloping land, at relatively low tree density, limited canopy size by pruning and bare soil management to optimize water use by the tree under the semiarid conditions which characterize the Mediterranean climate (Gómez, 2014). Indeed, there are many studies which

50 have measured high erosion rates in olive orchards on sloping areas (e.g. Gómez et al., 2014), although these high erosion rates are not necessarily a consequence of current management. Vanwallegghem et al. (2011) in a study of historical erosion rates in several ancient olive orchards of Montefrío (Southern Spain) reported unsustainable erosion rates in the range of 23 to  $68 \text{ Mg ha}^{-1} \text{ yr}^{-1}$  during the XIX and early XX centuries when these orchards were managed with bare soil, albeit based on animal ploughing. Vanwallegghem et al. (2011) also reported a further increase of these erosion rates when bare soil

55 management in these orchards started to be implemented in the late XX century using mechanization and herbicides. In the last five decades (Ruíz de Castroviejo, 1969), there has been an attempt to control soil degradation, while maintaining a favourable soil water balance for the tree through the gradual development of temporary cover crops (grown during the rainy season) (Gómez et al., 2014). These high erosion rates have also been linked to the degradation of soil properties observed in olive orchards. Gómez et al. (2009b) measured the differences in soil properties in a 5-year long experiment on runoff plots

60 reporting a decrease in SOC, aggregate stability and infiltration rates in bare soil management as compared to cover crops. Such scientific evidence which links changes of soil properties to different erosion rates in olive orchards under controlled conditions are rarely reported in the literature. Indeed, most of the studies connecting soil properties with different soil managements in olives come from surveys of soil properties in orchards on similar soil types but with differences in soil management. An example of these studies are those of Álvarez et al. (2010) or Soriano et al. (2014) who found an

65 improvement in soil properties, particularly aggregate stability, SOC and biological activity, in organic olive orchards with



cover crops, when compared to bare soil ones. In recent years, these studies have started to deepen our understanding in investigating key properties such as SOC. For instance, Vicente-Vicente et al. (2017) evaluated the impact of cover crops in the distribution of unprotected and protected SOC in the top 15 cm of the soil. These field studies take samples in a representative area of the slope, which is a common assumption in many soil quality studies (e.g. Andrews and Carroll, 2001). Although there are a limited number of experiments on the spatial variability of soil properties in olive orchards, they suggest a significant in-field variability (see Gargouri et al., 2013; Huang et al. 2017). Moreover, Gómez et al. (2012) suggested that part of this in-site variability of soil properties, regarding of organic carbon, might be related to erosion/deposition processes.

In-field variability associated with erosion/deposition processes is relatively well documented for organic carbon content in field crops (e.g., De Gryze et al. 2008, Mabit and Bernard, 1998, 2010; Van Oost et al., 2005). While the human-induced acceleration of soil erosion has depleted the SOC stock of agroecosystems, the fate of SOC transported over the landscape and that deposited in depressional sites is not fully understood, despite the fact that it might explain a high proportion of the in-site variability of soil properties.

Most of the erosion rates recorded or established in olive orchards come from runoff plots or small catchment experiments (e.g. Gómez et al., 2014). The use of the  $^{137}\text{Cs}$  approach has also demonstrated its potential in establishing long-term soil erosion rates in this specific land use. An example of these studies is that of Mabit et al. (2012) in which erosion as well as deposition rates since the 1950's were determined in one ancient olive orchard in the municipality of Montefrío, showing an average annual rate in the eroding part of the slope of  $12.3 \text{ t ha}^{-1} \text{ yr}^{-1}$ , and an average deposition rate in the lower section of the hillslope, much shorter than the eroding section, of  $13.1 \text{ t ha}^{-1} \text{ yr}^{-1}$ . This study involved a reference area for establishing precisely the initial  $^{137}\text{Cs}$  inventory, a natural undisturbed area located at 200 m from the orchard. To complement and/or to circumvent some limitation associated with the use of this anthropogenic radioisotope (see Mabit et al., 2008) and to maintain the capacity to determinate erosion and deposition rates without the need to use direct measurements, other natural radioisotopes such as  $^{210}\text{Pb}$  (e.g. Mabit et al., 2014; Matisoff et al., 2014) or stable isotopes such as  $\delta^{15}\text{N}$  or  $\delta^{13}\text{C}$  (e.g. Meusburger et al., 2013) have been proposed.

In this study, we hypothesized that the contribution of the long-term erosion-deposition processes on the in-field variability of soil properties in olive orchards (or other woody crops) under medium-high slope is relevant and should be taken into account when analysing the effects of specific strategies on SOC sequestration or on soil properties. In addition, we exploited the advantage provided by the unique location of an ancient olive orchard near an undisturbed reference area and the previous information on this site from studies on historical erosion rates, to fulfil the following objectives:

- 1- To quantify the long-term variability in soil total organic carbon and in their different fractions, and soil quality indicators in relation to erosion and deposition areas in an historical olive orchard;
- 2- To evaluate these differences in relation to the reference values found in an undisturbed natural area;
- 3- To evaluate differences in stable isotopes ( $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$ ) and explore their potential for identifying degraded areas within the olive orchard.



## 100 2 Materials and Methods

### 2.1 Description of the area

The study area is located in the municipality of Montefrío, southwestern Spain (Figure 1). The municipality extension is around 220 km<sup>2</sup>, of which 81 % is cultivated, mostly with olive trees. The climate in the region is continental Mediterranean with a long-term (1960–2018) average annual precipitation of 630 mm, a mean annual evapotranspiration of 750 mm, and a  
105 yearly average temperature of 15.2 °C. It is a mountainous area, with elevation ranging between 800–1600 m a.s.l at the highest point (Sierra de Parapanda). Soil sampling took place in two areas around the archaeological site “Peña de los Gitanos”, where the soils are classified as Calcic Cambisol according to the FAO classification. The reference undisturbed area was inside an archaeological site (Figure 1). This undisturbed area is covered by open Mediterranean forest interspersed with shrubs and annual grasses on limestone material (calcarenites). The status of this protected site guarantees that no  
110 anthropogenic activities have impacted it for a long period of time, approximately since the end of XVI century. Combined with its flat topography, this area has the potential to allow the establishment of reference values for undisturbed soil in the area. The area studied was an olive orchard located close to the reference area (Figure 1) which had been established in 1856. Both areas were described in detail in previous studies on historical erosion rates in the region (Vanwallegem et al., 2011, Mabit et al., 2012). This olive orchard is rainfed, and soil management in the decades before the sampling was based on bare  
115 soil with pruning residues (trees pruned every 2 years) being chopped and left on the soil surface. Olives are fertilized annually with 5 kg of 15 N-P-K per tree, spread below the tree canopy area.

### 2.2 Soil sampling

The reference site was sampled in two perpendicular transects, spaced at an average distance of 6 m. Using the excavation method, a total of 13 micropits in this reference site were collected per 5 cm increments until bedrock was reached (i.e. 0–5,  
120 5–10, 10–15, 15–20 and when possible, 20–25, 25–30, 30–35, 35–40, 40–45, 45–50, 50–55 and 55–60 cm). In the olive orchard a mechanical soil core of 8 cm diameter was used to sample 8 points in a 452 m long catena. At each sampling point soil was taken at different depths (0–10, 10–20, 20–30 and 30–40 cm). In a previous investigation, soil erosion and deposition rates were determined at each sampling point, comparing the <sup>137</sup>Cs inventory among these points and that of the undisturbed reference area (Mabit et al., 2012). The positions of all sampling points were recorded by RTK-GPS at submeter  
125 resolution (Table 1).

### 2.3 Physicochemical analysis

Soil samples were passed through a 2 mm sieve and homogenized, and stoniness determined as % in mass. Soil organic carbon concentration (C<sub>org</sub>) was determined according to Walkley and Black (1947). Separation of the various soil C<sub>org</sub> pools was performed by a combination of physical and chemical fractionation techniques through a three-step process developed  
130 by Six et al. (2002) and modified by Stewart et al. (2009), summarized here. First a partial dispersion and physical



fractionation of the soil is performed to obtain three size fractions: >250  $\mu\text{m}$  (coarse non-protected particulate organic matter, POM), 53–250  $\mu\text{m}$  (microaggregate fraction), and <53  $\mu\text{m}$  (easily dispersed silt and clay). This physical fractionation is done on air-dried 2-mm soil sieved over a 250- $\mu\text{m}$  sieve. Material greater than 250  $\mu\text{m}$  remained on the sieve. Microaggregates were collected on a 53- $\mu\text{m}$  sieve that was subsequently wet-sieved to separate the easily dispersed silt- and clay-sized fractions from the water-stable microaggregates. The suspension was centrifuged at 127 x g for 7 min to separate the silt-sized fraction. This supernatant was subsequently separated, flocculated and centrifuged at 1730 x g for 15 min to separate the clay-sized fraction. All fractions were dried in a 60 °C oven and weighed. Afterwards there is a second step involving a further fractionation of the microaggregate fraction isolated in the first step. A density flotation with Isodium polytungstate was used to isolate fine non-protected POM (LF): After removing the fine non-protected POM, the heavy fraction was dispersed overnight by shaking and passed through a 53  $\mu\text{m}$  sieve to separate the microaggregate-protected POM (>53  $\mu\text{m}$  in size, iPOM) from the microaggregate-derived silt and clay-sized fraction. The resulting suspension was centrifuged to separate the microaggregate-derived silt- versus clay-sized fraction as described above. A final third step involved the acid hydrolysis of each of the isolated silt- and clay-sized fractions. The silt- and clay-sized fractions from both the density flotation and the initial dispersion and physical fractionation were subjected to acid hydrolysis. The concentration of organic carbon in each of the isolated fraction was determined by wet oxidation using sulfuric acid and potassium dichromate following the methodology of Anderson and Ingram (1993). Inorganic carbon was removed prior to stable isotope analysis by acid fumigation following the method of Harris et al. (2001). Moistened subsamples were exposed to the exhalation of HCl in a desiccator overnight. Afterwards, the samples were dried at 40° C before measuring the stable isotope ratio. The N measurements were done with unacidified samples and the stable N isotope ratios and the C and N concentrations were measured by isotope ratio mass spectrometry (Isoprime 100 coupled with an Elementar Vario Isotope Select elemental analyser; both instruments supplied by Elementar, Langenselbold, Germany). The instrumental standard deviation for  $\delta^{15}\text{N}$  is 0.16% and 0.11% for  $\delta^{13}\text{C}$ . Stable isotopes are reported as delta values (‰) which are the relative differences between the isotope ratios of the samples and the isotope ratio of a reference standard.

In addition, available phosphorus ( $P_{\text{avail}}$ ) was determined by the Olsen method (Carter and Summers 1982) and organic nitrogen ( $N_{\text{org}}$ ) was determined by the Kjeldahl method (Stevenson, 1982). Water stable aggregates ( $W_{\text{sagg}}$ ) were measured using the method of Barthes and Roose (2002). Soil particle size distribution was determined using the hydrometer method (Bouyoucos, 1962) for the topsoil (0–10 cm) of the reference area and the olive orchard. As proposed by Hassink and Whitmore (1997), theoretical values of carbon saturation were established from the soil particle analysis. Finally, the soil degradation index developed by Gómez et al. (2009a) was calculated from the  $C_{\text{org}}$ ,  $P_{\text{avail}}$  and  $W_{\text{sagg}}$ .

## 2.4 Statistical analysis

The overall effect of depth and area (reference site vs. olive orchard or eroded vs. deposition area within the olive orchard) were evaluated using a two factor ANOVA ( $p < 0.05$ ). Additionally, for some comparison at similar soil depth, values of soil



properties between two different areas were assessed using a one-way ANOVA test ( $p < 0.05$ ) both situations, data were  
165 log-transformed when necessary to fulfil ANOVA requirements. The Pearson correlation coefficient was calculated among  
variables. The statistical software package Stata SE14.1 was used for these analyses.

### 3 Results



Table 2 shows the significance of the differences of bulk soil  $C_{org}$  and the various  $C_{org}$  fractions between reference and olive  
orchard plots, between soil depth and due to the interaction between both (Table 2A) and the effects of erosion/deposition  
170 ratio (Table 2B). Bulk soil  $C_{org}$  are always significantly higher in the reference area as compared to the olive orchard (Table  
2A and Figure 2A), and this is independent of the soil sampling depth.  $C_{org}$  values in the reference site are 2 to 5 times higher  
than that of the olive orchard for a given depth, with the greater differences in the top 10 cm of the soil. Soil depth has a  
significant effect on bulk  $C_{org}$  and  $C_{org}$  fractions, with values decreasing with depth in both areas. Unprotected, physically,  
175 chemically and biochemically protected fractions are higher in the reference site as compared to the olive orchard (Table 2A  
and Figure 3).  $C_{org}$  values are 2 to 6 times higher for the unprotected and chemically protected fractions, and between 2 to 3.5  
times higher for the physically and biochemically protected fractions, with differences tending to decrease with the soil  
depth.

Within the olive orchard, the overall analysis using a two-way ANOVA also highlights statistically significant differences  
between the erosion and deposition areas (Table 2B). Higher  $C_{org}$  values (1.1 to 0.6%) were observed in the deposition area  
180 located downslope, whereas lower values (0.85 to 0.55 %) were measured in the areas with net erosion in the upper and mid  
sections of the catena. It is worth noting that these differences between erosion and deposition areas are detected for the  
overall analysis using a two-way ANOVA (Tables 2A and B), although an individual analysis at each depth (Figure 2B) does  
not detect statistically significant differences, probably due to the moderate number of replications. Significant differences  
between the deposition and eroding area are also found for the unprotected and the physically and chemically protected  
185 fractions (Table 2B, Figure 4). However, differences for the biochemically protected (Table 2B, Figure 4) are not.

The percentage distribution of SOC fractions is also significantly different between both areas (reference vs. olive orchard),  
except for the biochemically protected fraction (Table 3A, Figure 5). The reference area stores most of the  $C_{org}$  in the  
unprotected fraction (between 50 and 65% approximately) with no significant trend with depth (Table 3A, Figure 5),  
followed in relative importance by the chemically and physically protected fractions which contain between 18–30 % and  
190 10–20 % of the bulk soil  $C_{org}$ , respectively. The biochemically protected fraction represents a very low percentage (between  
4 to 6 %). In the olive orchard,  $C_{org}$  is stored predominantly in the physically and chemically protected fractions which  
accounted for about 38 to 27 and 34 to 28 % respectively, followed by the pool of unprotected fraction (between the 22 to  
32%) (Figure 5).



The biochemically protected fraction represents from 11 to 4% of the organic carbon stored in the olive orchard, approximately. There are no clear differences in the organic carbon distribution among the different fractions between the erosion and deposition areas in the olive orchard (Table 3B and Figure 6).

Figure 7 shows the mean SOC stock for the top 40 cm of the soil. SOC stock in the reference site is as large as 120 t ha<sup>-1</sup> and is significantly higher than the olive orchard which stores between 41 and 54 t ha<sup>-1</sup> in the eroded and deposition areas respectively, without significant differences between these two.

Texture distribution of the topsoil (0–10 cm) along the catena in the olive orchard presents an average clay, silt and sand content of 41, 37 and 22% and low variability, respectively (average coefficient of variation of 17%) without significant changes between the erosion and deposition areas. In the reference area, the soil has an average clay, silt and sand content of 30, 31 and 39% respectively, also with a homogeneous distribution across the sampling area (coefficient of variation of 10%). According to the Hassink and Whitmore (1997) model, the percentages of organic carbon of maximum soil stable C<sub>org</sub> are of 1.94 and 1.15 % in the reference site and olive orchard, respectively. So, protected C<sub>org</sub> in the reference and olive orchard areas account for 87 % and 64 % of the maximum soil stable C<sub>org</sub>, respectively at the topsoil

Figure 8 and Table 4A compare stable isotope delta values between the reference site and the overall olive orchard by depth. There are statistically significant differences in  $\delta^{15}\text{N}$ ,  $\delta^{13}\text{C}$ , and  $\delta^{13}\text{C}:\delta^{15}\text{N}$  ratio between the two areas, although in the case of  $\delta^{15}\text{N}$  at the top 20 cm, soil depth had a significant effect. When comparing differences between the erosion and deposition areas within the olive orchard, we detected statistically significant differences only in  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}:\delta^{15}\text{N}$  ratio, especially in the 10–20 cm of the soil (Figure 9, Table 4B).

Figure 10 depicts the comparison between the P<sub>avail</sub>, N<sub>org</sub>, W<sub>sagg</sub> as well as the soil degradation index (SDI, Gómez et al. 2009a) at the top 10 cm of the soil between the erosion and deposition area of the olive orchard. Table 5 presents a similar comparison for N<sub>org</sub>, P<sub>avail</sub> and bulk density at the different soil depths. P<sub>avail</sub> in the deposition area is much higher than that of the erosion area, with a similar trend, while not statistically significant, for N<sub>org</sub> and W<sub>sagg</sub>. SDI, which is an aggregated indicator of these three soil variables, in the eroded area is about 3 times higher than that in the deposition area.

#### 4 Discussion

After approximately 175 years of contrasted land use between the undisturbed reference site and the olive orchard, bulk soil organic carbon concentration and its fractions have been dramatically reduced in the olive orchard. Current levels of C<sub>org</sub> concentration in the soil profile are approximately 20–25% of the reference area covered by the natural vegetation in the area adjacent to the orchard. This ratio is similar, albeit in the lower range, of the comparison of C<sub>org</sub> in topsoil among olive orchards with different managements and natural areas reported for the region (Millgroom et al., 2007). The increased soil disturbance, the lower annual rate of biomass returned to the soil and the higher erosion rate in the olive orchard explain this difference. In both areas, the C<sub>org</sub> is clearly stratified, indicating that despite the different mechanisms involved there is a periodic input of biomass from the olive trees (e.g. fall down of senescence leaves and tree pruning residues) plus the annual



ground vegetation. Vicente-Vicente et al. (2017) estimated this biomass contribution in the range of 1.48 to 0.56 t ha<sup>-1</sup> yr<sup>-1</sup>. It is worth noticing that the decrease in C<sub>org</sub> as compared to the natural area is much higher than the reported rates of increase in C<sub>org</sub> in olive orchards using conservation agriculture (CA) techniques, such as cover crops and incorporation of organic residues from different sources. In a meta-analysis Vicente-Vicente et al. (2016) found a response ratio (the ratio of C<sub>org</sub> under CA management as compared to bare soil managed orchard) from 1.1 to 1.9 suggesting that under a CA management, which combines cover crops and organic residues, C<sub>org</sub> doubled as a maximum.

Combining all C<sub>org</sub> data of the olive orchard, the variability was about 35% which is similar to what has been reported so far in the few studies on soil C<sub>org</sub> variability in olive orchards. For instance, Gargouri et al. (2013) indicated a 24% coefficient of variation (CV) in a 34 ha olive orchard in Tunisia, while Huang et al. (2017) reported an average CV of 41% in a 6.2 ha olive orchard in Southern Spain. Neither of these two studies reported clear trends on the distribution of C<sub>org</sub> with topography. Huang et al. (2017) pointed out the additional difficulties in the determination of C<sub>org</sub> due to the topography heterogeneity, although this was compounded by the fact that within the orchards there were two areas with different planting dates for the trees. Gómez et al. (2012) reported a CV of 49% with higher C<sub>org</sub> in areas where there was a change in slope gradient from the hillslope to a draining central channel into the catchment, although they could not find a simple relationship between the increase in content and the topographic indexes. Despite the fact that a lot of work has been done on the correlation between erosion-deposition and the redistribution of soil C<sub>org</sub>, (e.g. Van Oost et al. (2005)), our study is, to our knowledge, the first attempt to quantify this in detail under olive orchard agro-environmental condition. The variability induced by the combined effects of water and tillage erosion in this olive orchard was similar to that described in other agroecosystems. For instance, Van Oost et al. (2005) measured on two field crop sites under temperate climate, a clear correlation between the erosion-deposition rates and the topsoil C<sub>org</sub> concentration, which ranged between 0.8 % of the erosion to 1.4% of the deposition sites in the top 25 cm of the soil. Besides this, Bameri et al. (2015) in a field crop site with a semi-arid environment, measured also a higher C<sub>org</sub> in lower part of the field where deposition of the eroded soil from the upper zones took place with a mean C<sub>org</sub> value of 0.95% in the top 20 cm of the soil and a CV of 53%. Overall, under such landscapes cultivated for a long time, the cumulative effect of tillage and water erosion on the redistribution of soil across the slope has been observed (Dlugoß et al., 2012). These processes also produce a vertical redistribution of C<sub>org</sub> resulting in a relatively homogeneous profile in the tilled layer (top 15–20 cm) and a gradual decline below this depth, as noted in this study.

This horizontal distribution due to tillage and water erosion also simultaneously affected other soil properties and has been described previously in other field crops areas. For instance, De Gryze et al. (2008) described, in a field crop area under conventional tillage in Belgium, how P<sub>avail</sub> almost doubled (22.9 vs. 12.2 mg kg<sup>-1</sup>) in the depositional area as compared to the eroding upper part. They also reported that in half of the field under conservation tillage these differences in P<sub>avail</sub> between the upper and lower areas of the field disappeared. We have also observed in our sampled orchard a pronounced increase in topsoil P<sub>avail</sub> in the deposition area of around 400% as compared to the eroding part of the orchard. The cumulative effects of the differences in C<sub>org</sub>, P<sub>avail</sub>, and the trend towards higher, although non-significant, W<sub>sagg</sub> in the deposition area, allow us to



260 delimit two distinctive areas within the orchard with marked differences in soil quality: the eroded part which is within the  
range considered as degraded in the region (Gómez et al., (2009a) and the depositional area which is within the range of non-  
degraded according to the same index, and represents the 20% of the orchard transect length (Table 1). This again raises the  
need for a careful delineation of sub-areas when analysing soil quality indicators and/or SOC carbon stock within the same  
field unit. Topography and sediment redistribution by erosive processes introduce a gradient of spatial variability that  
265 questions the concept of representative area when it comes to describing a whole field. In fact, several studies (e.g. Dell and  
Sharpley, 2006), have suggested that the verification of compliance of environmental programs such as those related to  $C_{org}$   
sequestration should be based preferentially, at least partially, on modelling approaches.

The differences between the reference site and the olive orchard are similar to those described previously when comparing  
cropland and forested areas, with the latter presenting a higher concentration of  $C_{org}$  (and most of it in the unprotected  
270 fraction) while the cropland presented a higher fraction of the carbon in the physically and chemically protected fractions  
(e.g. Poeplau and Don, 2013). This is likely due to the fact that under soil degradation and low annual organic carbon inputs,  
as is the case under olive orchard land use, most of the unprotected  $C_{org}$  decomposes relatively quickly and a great proportion  
of the remained low SOC is protected. In addition, the mobilisation of the unprotected  $C_{org}$  is expected to be reduced in the  
protected forested area because of the canopy and the existing vegetation on the ground that protects the soil against runoff  
275 and splash erosion processes. In fact, the protected  $C_{org}$  concentration in the topsoil of the olive orchard in the eroded area is  
about the 60 % of the upper limit of protected  $C_{org}$  (1.19 %) according to the model of Hassink and Whitmore (1997).  
Therefore, the low unprotected SOC concentration found in the olive orchard is an issue in the increase of SOC stocks.  
Furthermore, the percentage of organic carbon of maximum soil stable  $C_{org}$  in the olive orchard was 59 % of that of the  
reference site, and therefore the soil degradation in the olive orchard does not only decrease the level of  $C_{org}$  but also the  
280 capacity for  $C_{org}$  stabilization. This is because protected fractions are fuelled from recently derived, partially decomposed  
plant residues together with microbial and micro, meso and macrofaunal debris (unprotected organic carbon) throughout  
processes like SOC aggregation into macro- and/or microaggregates (physically protected SOC) and complex SOC  
associations with clay and silt particles (chemically protected SOC) which are disrupted in the cropland area as in  
comparison with the reference area. The distribution among soil  $C_{org}$  fractions in the orchard of this study was similar to the  
285 result obtained by Vicente-Vicente et al. (2017) who measured  $C_{org}$  fractions distribution in olive oil orchards with  
temporary cover crops, with the exception of the unprotected SOC, which was much higher in soils under cover crops than  
that of our study under bare soils. The study of Vicente-Vicente et al. (2017) showed a large variability among orchards  
attributed, among other reasons, to the large variability in biomass production by the cover crops in the orchards. In our  
study, the erosion/deposition processes also induced significant differences in  $C_{org}$  concentration and contributed to  
290 increasing the variability in the distribution of organic carbon among the different areas of the olive orchard. The greatest  
differences between eroded and deposited area were observed for the unprotected SOC, especially for the top 10 cm of soils.  
This is likely due to the transport of the low density, and prone to water floating, of the partially decomposed plant and  
microbial and animal residues from the eroded part to the deposited part of the orchard. Differences in physically and



chemically protected SOC between eroded and deposited areas also indicate selective deposition of soil aggregates, although  
295 this is not reflected in the change in topsoil texture, which could be explained by the homogenizing effect of other processes  
of soil redistribution across the slope (e.g. tillage erosion, creeping, ...) and the relatively high silt and clay content of the  
soil.

The relatively high variability of  $C_{org}$  and other soil quality indicators due to erosion/deposition process indicates that this  
variability needs to be addressed for a proper appraisal of the distribution of organic carbon among the different fractions in  
300 a given orchard. Differences in vegetation types induced differences in  $\delta^{13}C$  between the olive orchard and the reference  
area but, as expected, no differences in  $\delta^{13}C$  were detected between the erosion and deposition areas in the olive orchard  
given the same origin of vegetation derived organic matter, C3-plants. Interestingly, within the olive orchard, significant  
differences in  $\delta^{15}N$  were detected between the erosion and deposition areas, especially for the top 20 cm of soils (Figure 9).  
This suggests the possibility of using  $\delta^{15}N$  as a variable for identifying degraded area in olive growing fields, as has been  
305 proposed for other eroding regions in the world (e.g. Meusburger et al. 2013), which might provide an alternative when other  
conventional or isotopic techniques are not available. The source of N in soil is multifarious and subject to a wide range of  
transformations that affect  $\delta^{15}N$  signature and therefore we can only speculate on the reasons for this difference in  $\delta^{15}N$  in a  
relatively homogeneous area. Bulk soil  $\delta^{15}N$  tended to be more positive (e.g. more enriched in  $\delta^{15}N$ ) as N cycling rate  
increases soil microbial processes (e.g. N mineralization, nitrification and denitrification) resulting in products (e.g. nitrate,  
310  $N_2O$ ,  $N_2$ ,  $NH_3$ ) depleted in  $^{15}N$  while the substrate from which they were formed becomes slightly enriched (Robison, 2001).  
The higher  $\delta^{15}N$  signature of the soil at deposition location suggests that rates of processes involved in the N cycling are  
higher than in the erosion area and that is in accordance with the higher bulk  $C_{org}$  and  $P_{avail}$  contents and lower SDI of the  
deposition site. The relatively lower soil  $\delta^{15}N$  signature at the reference site could be partially due to the input of litter N  
from the natural legumes and to the closed N cycling which characterize natural forest ecosystems. The trend in  $^{15}N$   
315 enrichment with soil depth, as found in the reference site, is a common observation in forest and grassland sites, which has  
been related to different mechanisms, including  $^{15}N$  isotope discrimination during microbial N transformations, differential  
preservation of  $^{15}N$ -enriched soil organic matter components during N decomposition, and more recently, to the build-up of  
microbial  $^{15}N$ -enriched microbial necromass (Huygens et al., 2008). However, there still remains the need for a careful  
calibration against an undisturbed reference site and a better understanding of the influence of different vegetation between  
320 the reference and the studied area in the change of the  $\delta^{15}N$  signal for its further use as an additional tool to determine soil  
degradation.

## 5 Conclusions

1- The results indicate that erosion and deposition within the investigated old olive orchard have created a significant  
difference in soil properties along a catena, which is translated into different soil  $C_{org}$ ,  $P_{avail}$  and  $N_{org}$  contents and  $\delta^{15}N$ , and  
325 thus soil quality status.



- 2- This variability was smaller than that of the natural area, which indicated a severe depletion of SOC as compared to the natural area and a redistribution of available organic carbon among the different SOC fractions.
- 3- The results suggest that  $\delta^{15}\text{N}$  has the potential for been used as an indicator of soil degradation. More investigation under different agroecosystems would be required for confirming this statement at larger scale.
- 330 4- This investigation highlights that proper understanding and management of soil quality and  $C_{\text{org}}$  content in olive orchards require considering the on-site spatial variability induced by soil erosion/deposition processes.

## 6 Acknowledgements

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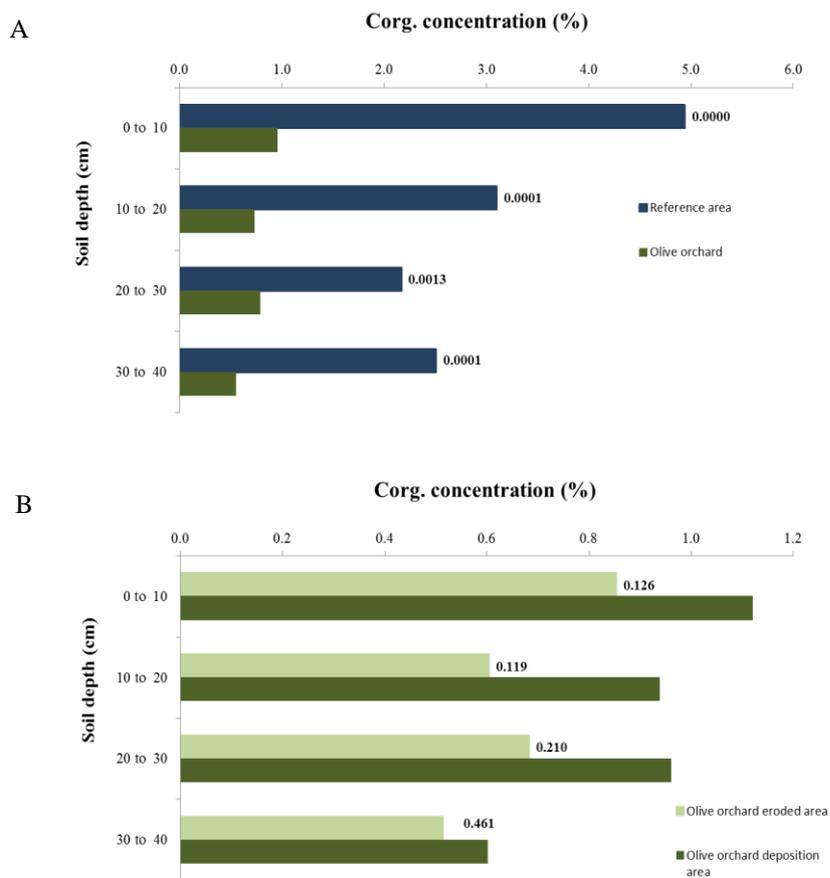


460 **Figure 1: Site location and associated sampling. Left: Location map of the sampling area in Montefrío, Southern Spain. Reference**  
**site limited by the white line within a protected archaeological site (yellow line). Yellow arrows indicate the sampled transect**  
**within the olive orchard. Right: Location of the sampled points in transect at the olive orchard. Numbering starts in the points at**  
**higher elevation. Europe map designed by Freepik, and air images source Google Earth (© Google 2018).**





475 **Figure 2: Comparison of average soil organic carbon concentration in bulk soil by soil depth between: A) reference site vs. olive**  
**orchard; B) eroded and deposition areas within the olive orchard. Labels in bars indicate the p-value according to a one-way**  
**ANOVA between treatments for the same soil depth.**



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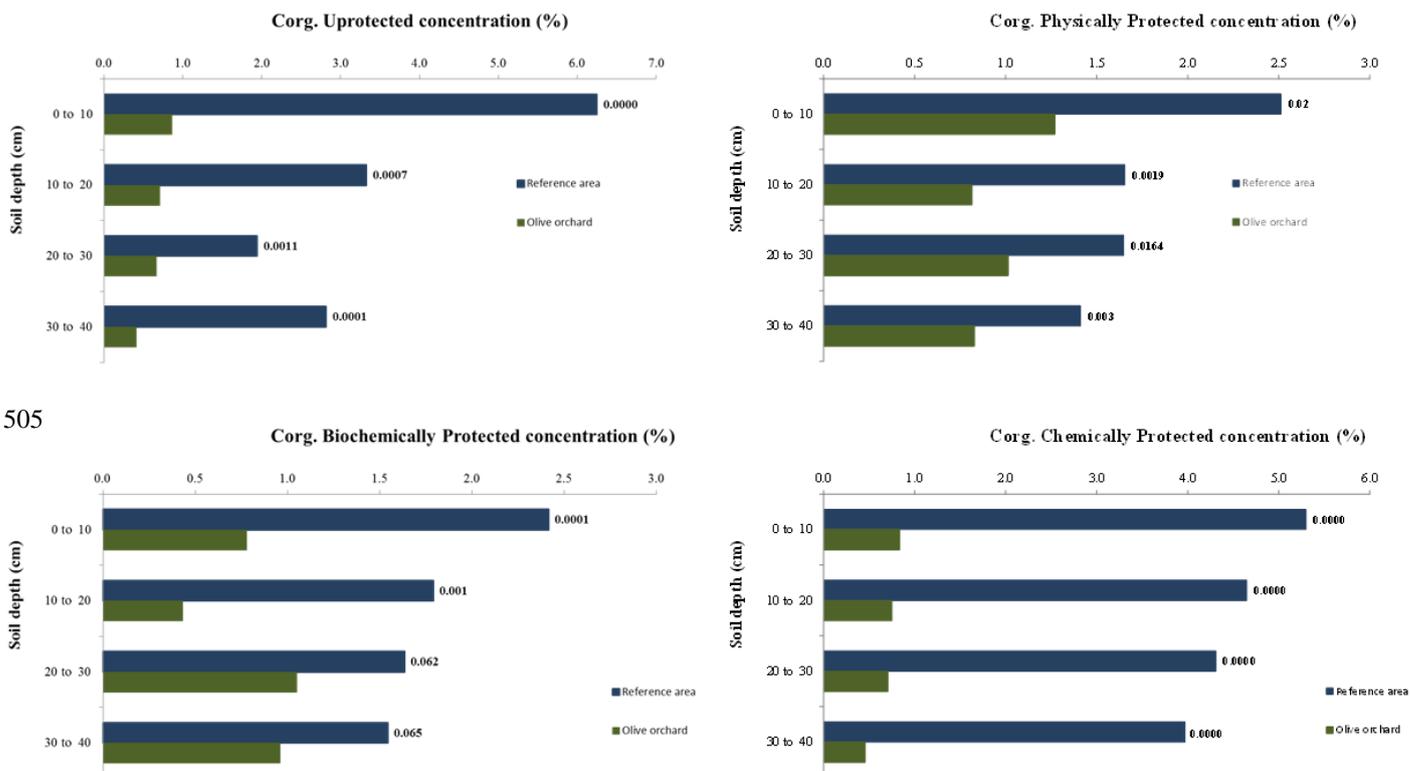
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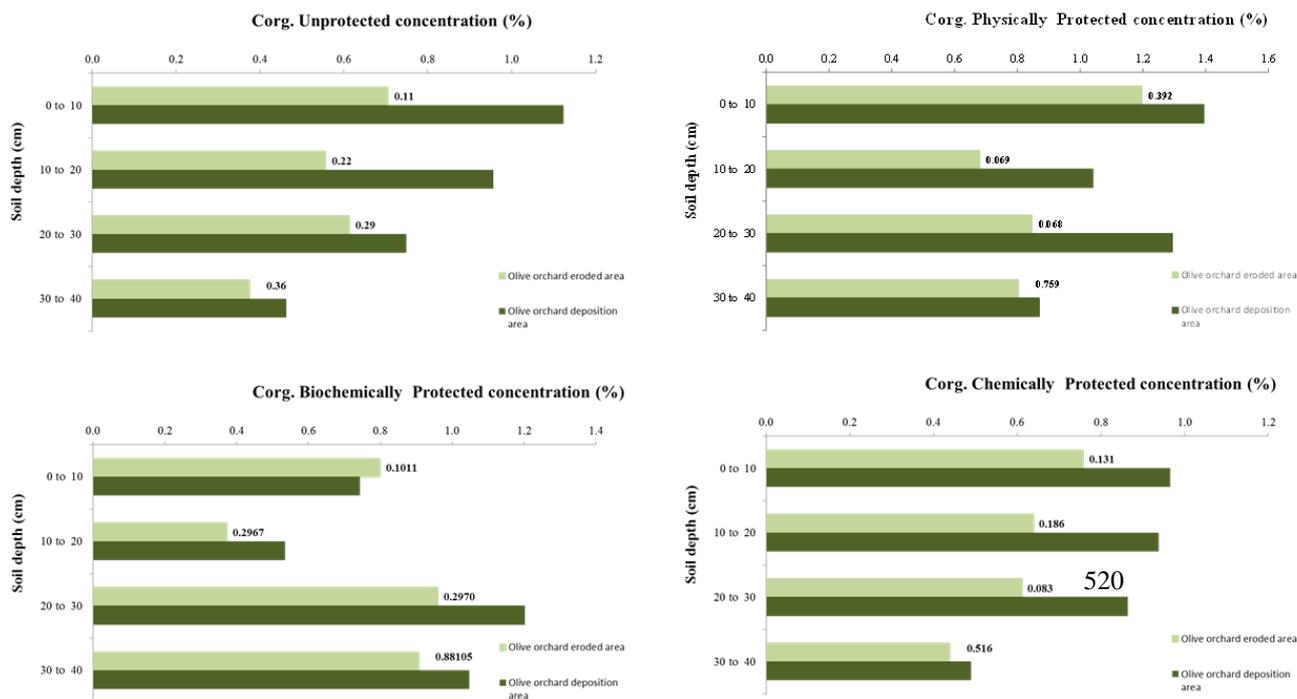
**Figure 3: Organic carbon concentration in the different soil organic carbon fractions at each depth comparing reference site vs. olive orchard. Labels in bars indicate the p-value according to a one-way ANOVA comparing treatments for the same soil depth.**



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510 **Figure 4: Organic carbon concentration in the different soil organic carbon fractions by depth comparing erosion vs. deposition areas within the olive orchard. Labels in bars indicate the p-value according to a one-way ANOVA comparing areas for the same soil depth.**

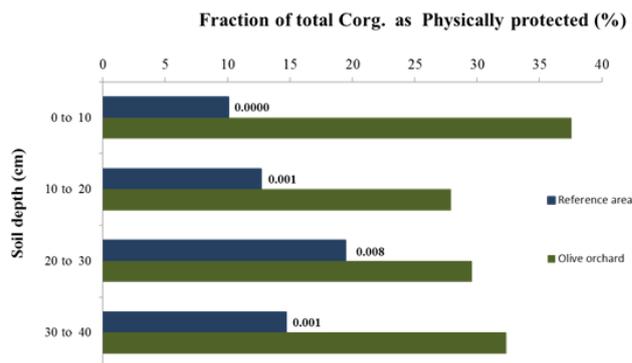
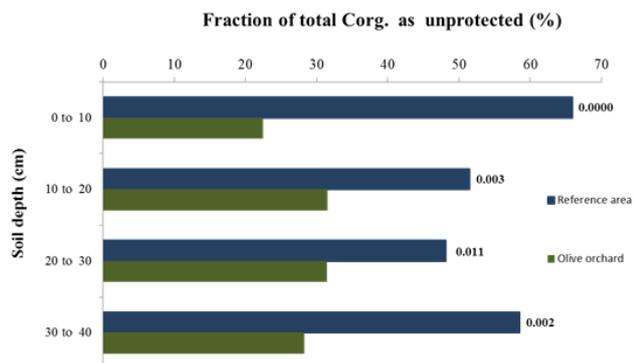




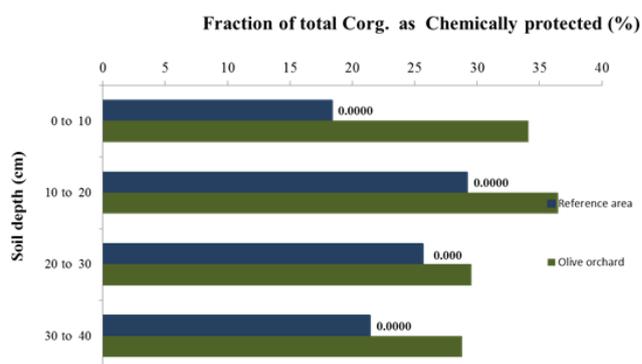
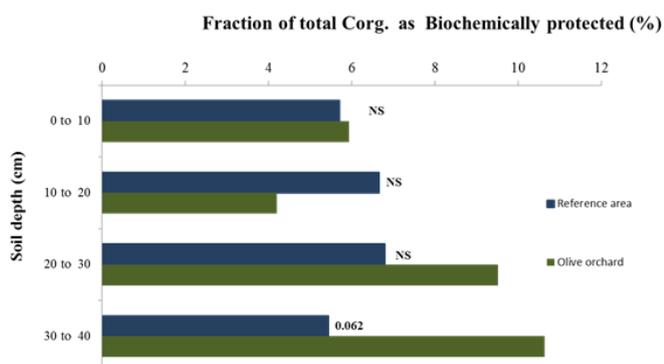
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**Figure 5: Fraction of total organic carbon stored in the different fractions by depth comparing reference site vs. olive orchard. Labels in bars indicate the p-value according to a one-way ANOVA comparing areas for the same soil depth.**

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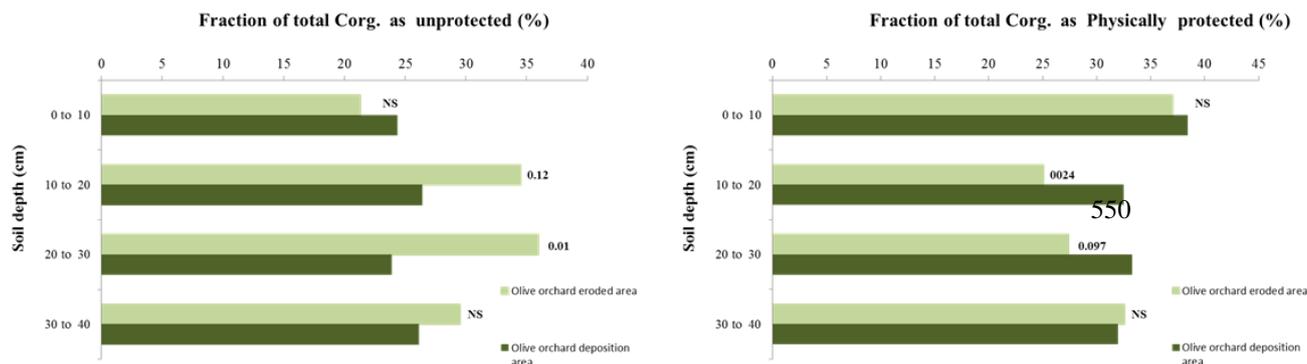


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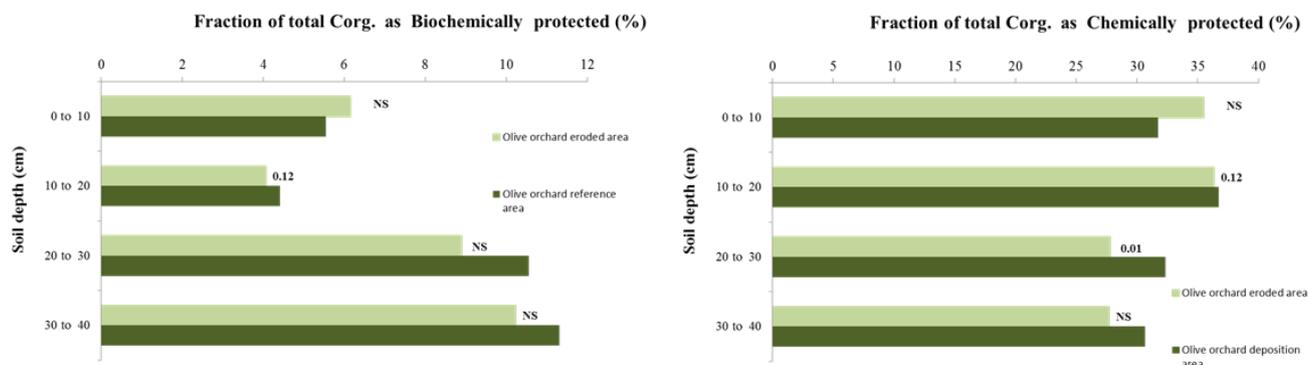


**Figure 6: Fraction of total organic carbon stored in the different fractions by depth comparing erosion vs. deposition areas within the olive orchard. Labels in bars indicate the p-value according to a one-way ANOVA comparing areas for the same soil depth.**

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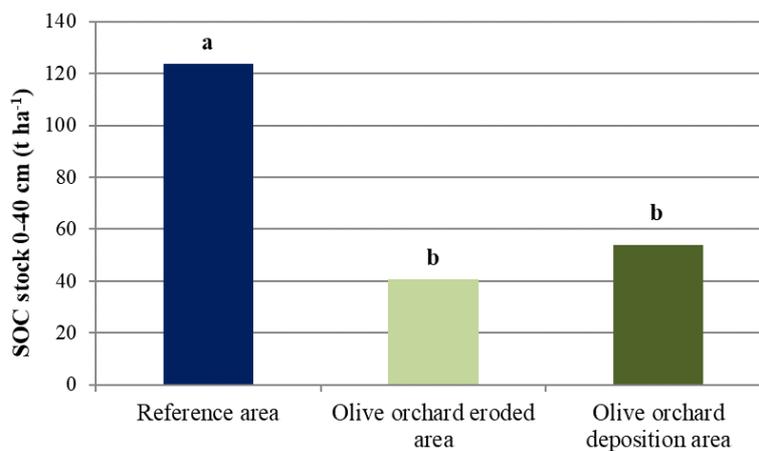
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**Figure 7: Total soil organic carbon (SOC) stock at the top 40 cm of the soil in the reference site and in the erosion and deposition areas of the olive orchard. Different letters above bars means statistically significant differences (Kruskal-Wallis test at  $p < 0.05$ ).**

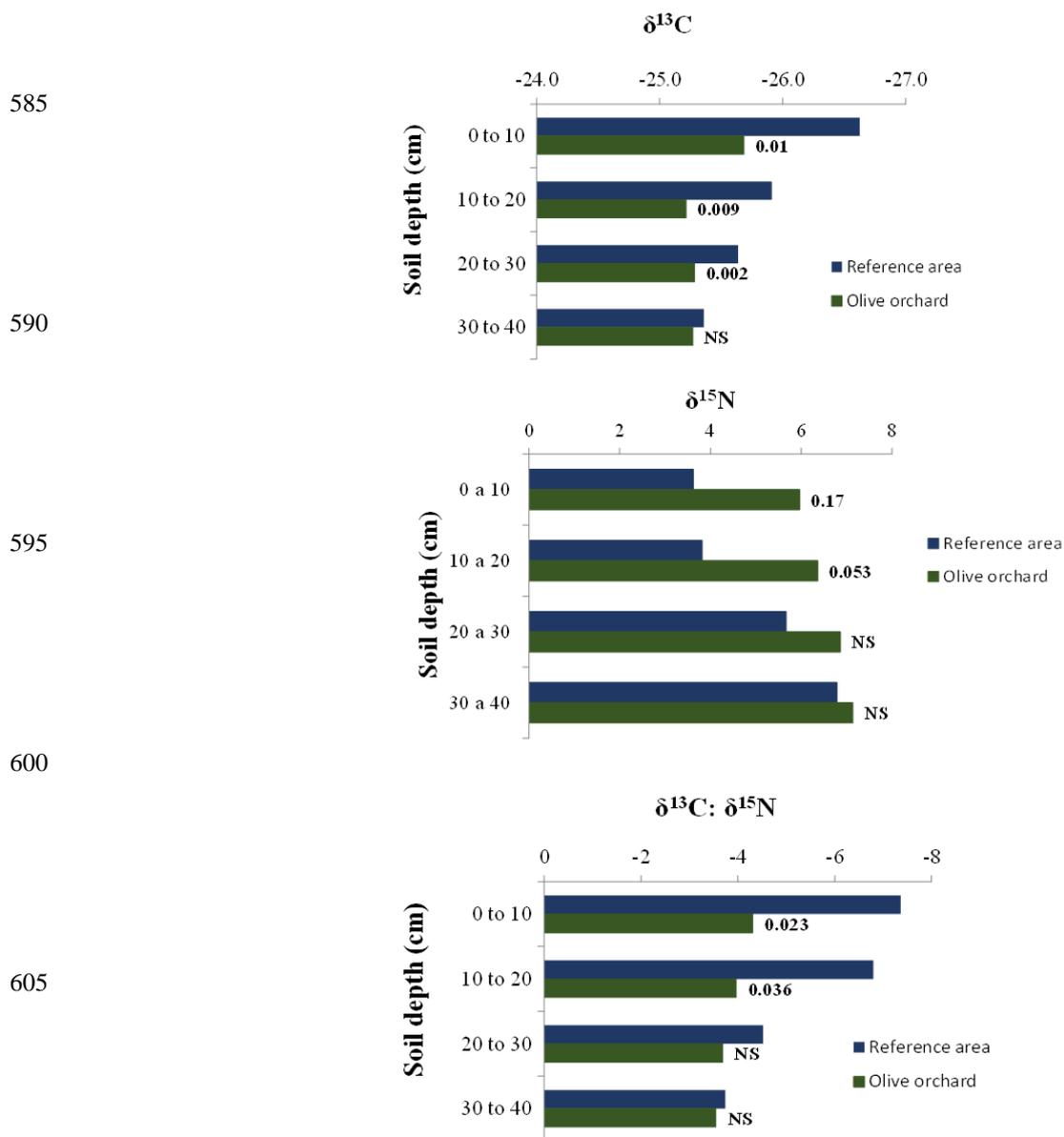
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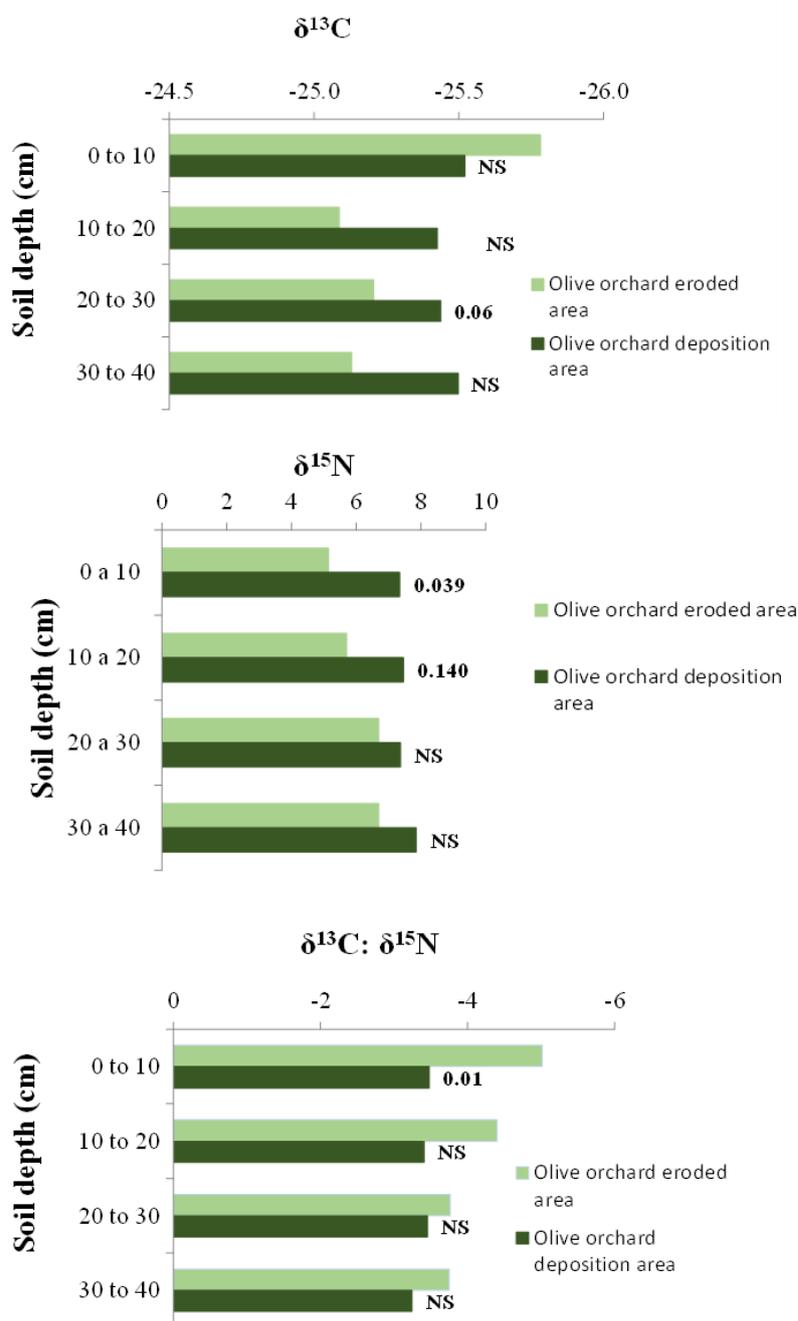


580 **Figure 8:**  $^{13}\text{C}$  and  $^{15}\text{N}$  isotopic signal of soil by depth comparing reference site vs. olive orchard. Labels in bars indicate the p-value  
according to a one-way ANOVA comparing treatments for the same soil depth.





**Figure 9:**  $^{13}\text{C}$  and  $^{15}\text{N}$  isotopic signal of soil by depth comparing erosion vs. deposition areas within the olive orchard. Labels in bars indicate the p-value according to a one-way ANOVA comparing area treatments for the same soil depth.

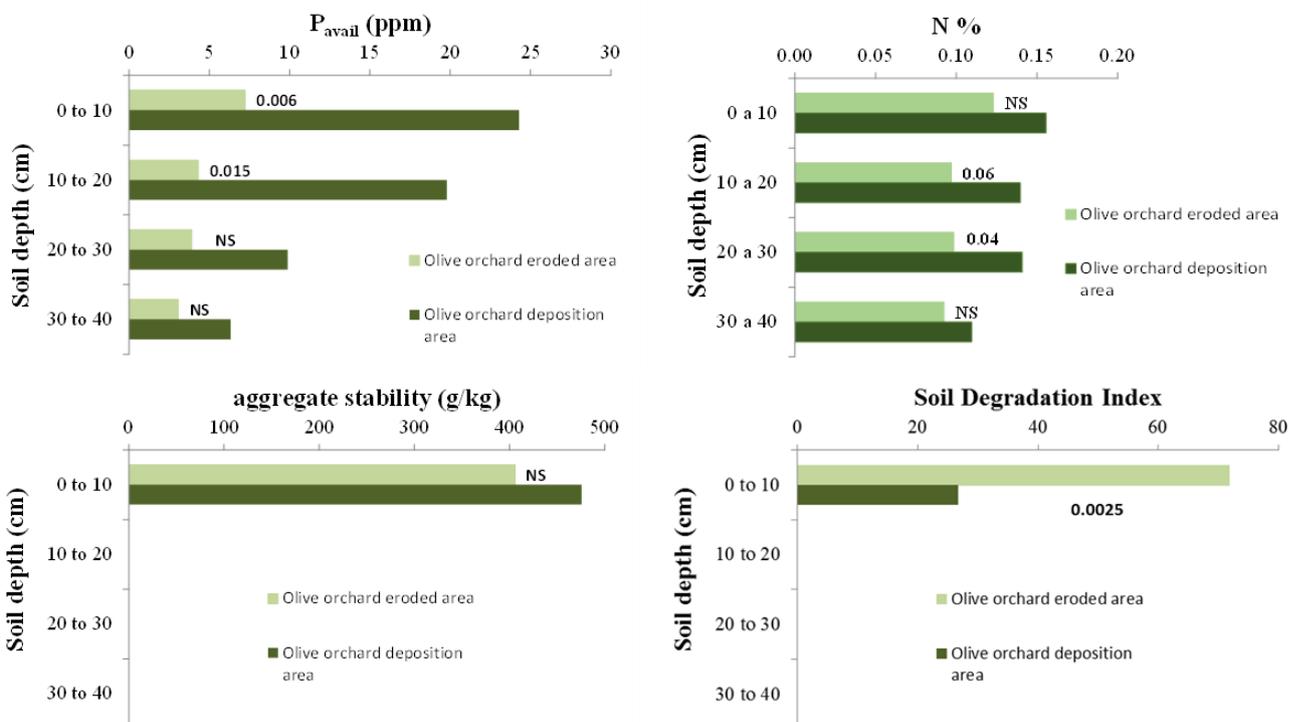




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**Figure 10: Soil available phosphorus ( $P_{avail}$ ), organic nitrogen ( $N_{org}$ ), aggregate stability ( $W_{sagg}$ ) and Soil Degradation Index (SDI) by depth comparing eroded vs. deposition areas within the olive orchard. Labels in bars indicate the p-value according to a one-way ANOVA comparing areas for the same soil depth.**

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670 **Table 1: Location of the sampling points along the transect and associated soil redistribution rates derived from the**  
**<sup>137</sup>Cs technique (adapted from Mabit et al., 2012). Negative values indicate net erosion and positive values net**  
**deposition.**



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Point #	Code	Distance in transect (m)	Elevation (m)	Erosion/deposition rate (t ha <sup>-1</sup> yr <sup>-1</sup> )
1	Cs1	0	1044	-5.2
2	Cs3	66.4	1032.8	-17.8
3	Cs5	125.0	1017.8	-7.1
4	Cs7	179.0	1006.8	-16.3
5	Cs12	312.2	986.8	-15.2
6	Cs13	338.1	984.8	5.9
7	Cs15	388.8	981.8	24.7
8	Cs17	429.5	979.8	8.8



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**Table 2: Results of the two-way ANOVA analysis of soil organic carbon concentration,  $C_{org}$ , in different fractions and in bulk soil. In A) area refers to reference site vs. olive orchard and in B) area refers to eroded vs. deposition areas in the olive orchard. NS stands for Not Significant.**

690 A)

Model	Bulk soil	$C_{org}$ fraction			
		Not protected	Physically protected	Chemically protected	Biochemically protected
Area (A)	0.0000	0.0000	0.0000	0.0000	0.0000
Depth (D)	0.0023	0.0022	0.0061	0.0190	NS
A x D	NS	NS	NS	NS	0.0300

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700 B)

Model	Bulk soil	$C_{org}$ fraction			
		Not protected	Physically protected	Chemically protected	Biochemically protected
Area (A)	0.0198	0.0400	0.0077	0.0299	NS
Depth (D)	0.0081	0.0070	0.0058	0.0055	0.0847
A x D	NS	NS	NS	NS	NS

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**Table 3: Results of the two-way ANOVA analysis of the distribution of the total soil organic carbon content in the soil among the different fractions of soil organic carbon,  $C_{org}$ . In A) area refers to reference site vs. olive orchard and in B) area refers to eroded vs. deposition areas in the olive orchard. NS stands for Not Significant.**

715 A)

Model	$C_{org}$ fraction			
	Not protected	Physically protected	Chemically protected	Biochemically protected
Area (A)	0.0000	0.0000	0.0000	NS
Depth (D)	NS	NS	0.0640	NS
A x D	NS	0.0059	NS	NS

720

B)

Model	$C_{org}$ fraction			
	Not protected	Physically protected	Chemically protected	Biochemically protected
Area (A)	0.091	0.0881	NS	NS
Depth (D)	0.051	0.0214	NS	0.033
A x D	NS	NS	NS	NS

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**Table 4: Results of the two-way ANOVA analysis of the stable isotopes signal. In A) area refers to reference site vs. olive orchard and in B) area refers to eroded vs. deposition areas in the olive orchard. NS stands for Not Significant.**

A)

735

Model	$\delta^{13}\text{C}$	$\delta^{15}\text{N}$	$\delta^{13}\text{C} : \delta^{15}\text{N}$
Area (A)	0.0001	0.002	0.002
Depth (D)	0.0026	0.0175	0.029
A x D	NS	NS	NS

B)

740

Model	$\delta^{13}\text{C}$	$\delta^{15}\text{N}$	$\delta^{13}\text{C} : \delta^{15}\text{N}$
Area (A)	NS	0.01	0.01
Depth (D)	NS	NS	NS
A x D	NS	NS	NS

745



**Table 5: Results of the two-way ANOVA analysis of some soil physical and chemical properties comparing eroded vs. deposition areas in the olive orchard. NS stands for Not Significant.**

750

<b>Model</b>	<b>N<sub>org</sub></b>	<b>P<sub>avail</sub></b>	<b>Bulk density</b>
Area (A)	0.0000	0.01	NS
Depth (D)	0.0009	NS	NS
A x D	NS	NS	NS

755